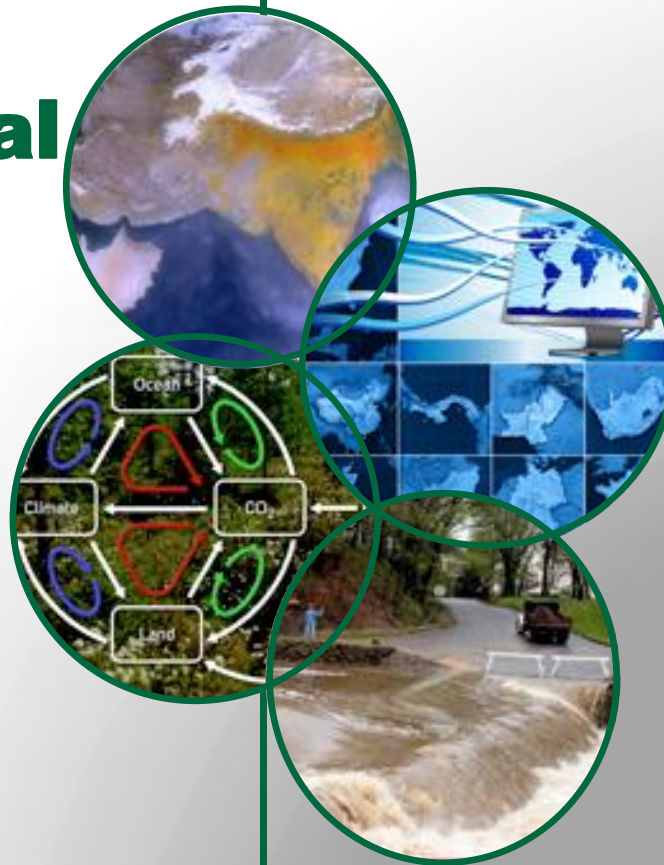


# Temperature response of enzyme activity at depth in a bog at Marcell Experimental Forest, MN, USA

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**<sup>3</sup>University of Tennessee**



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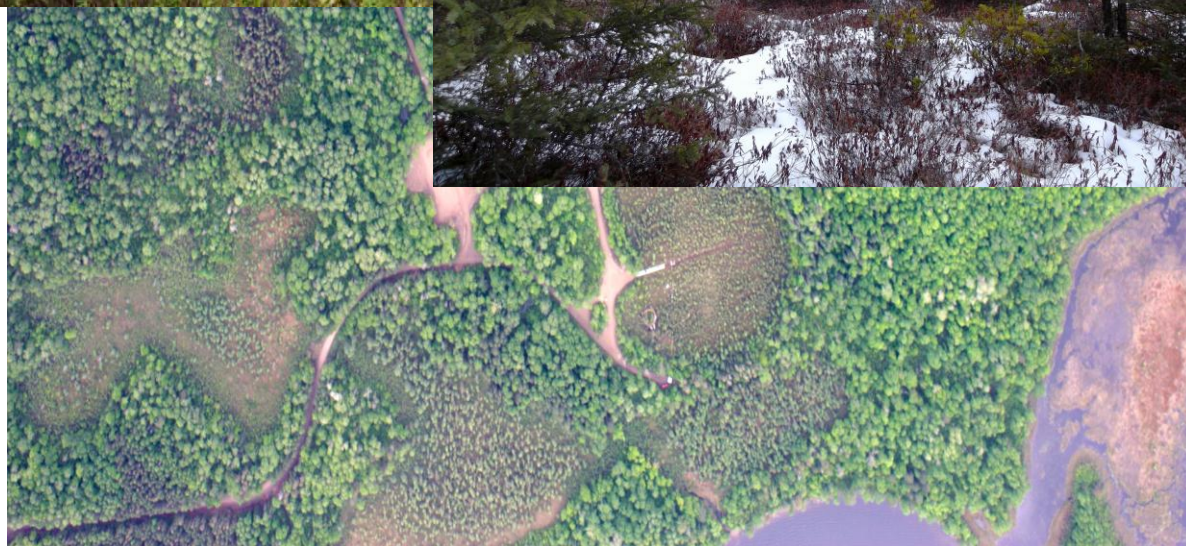
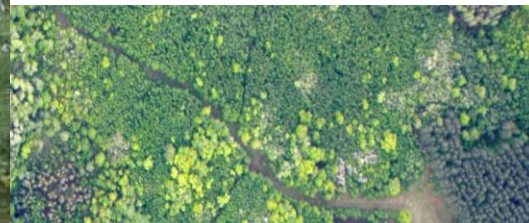


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# introduction: SPRUCE

Spruce and  
Peatland  
Responses  
Under  
Climate and  
Environmental change



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# introduction: SPRUCE design

- Warming – above and belowground
  - 0-9° C above ambient
- CO<sub>2</sub>
  - ambient and elevated

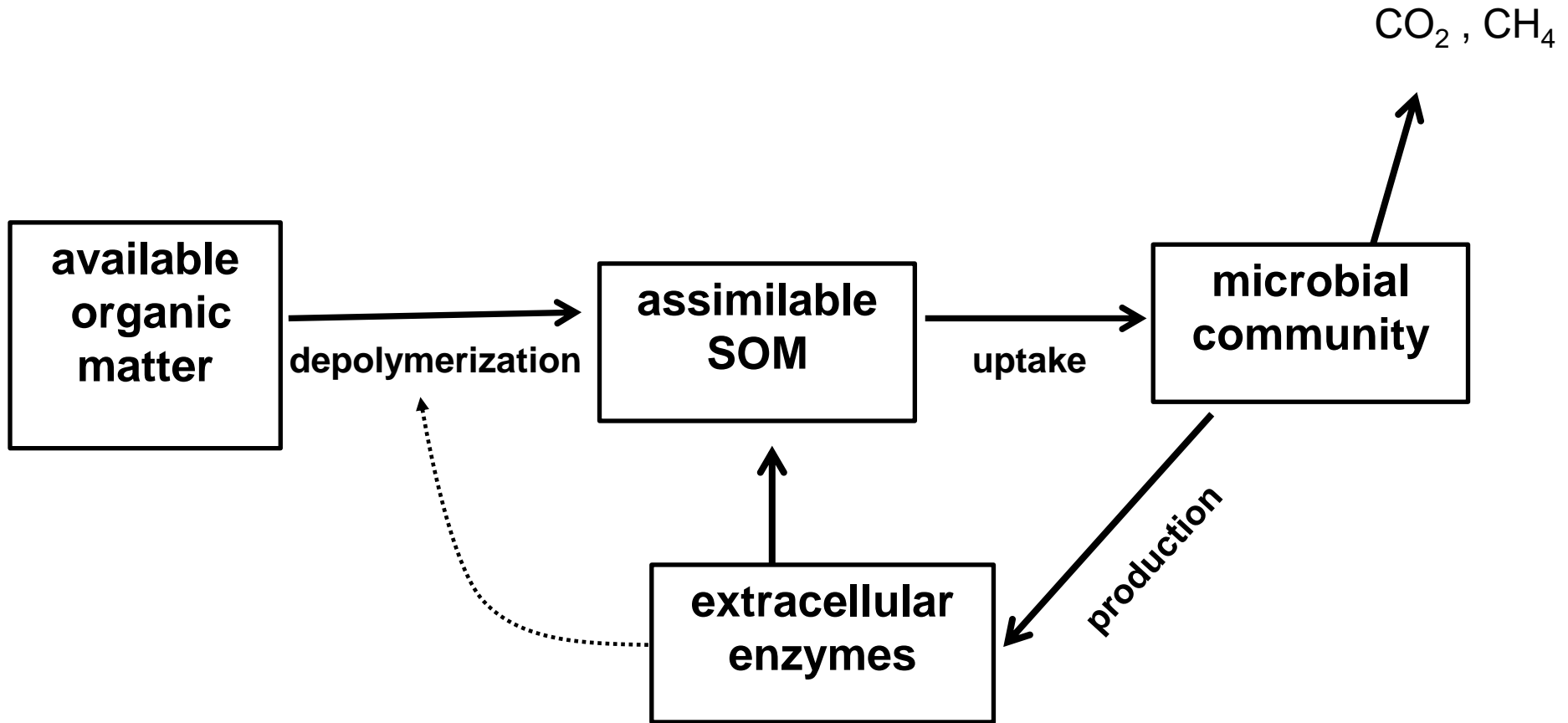


Full-scale prototype of the enclosure for testing evaluation and continued improvement constructed at ORNL.

## Information about SPRUCE

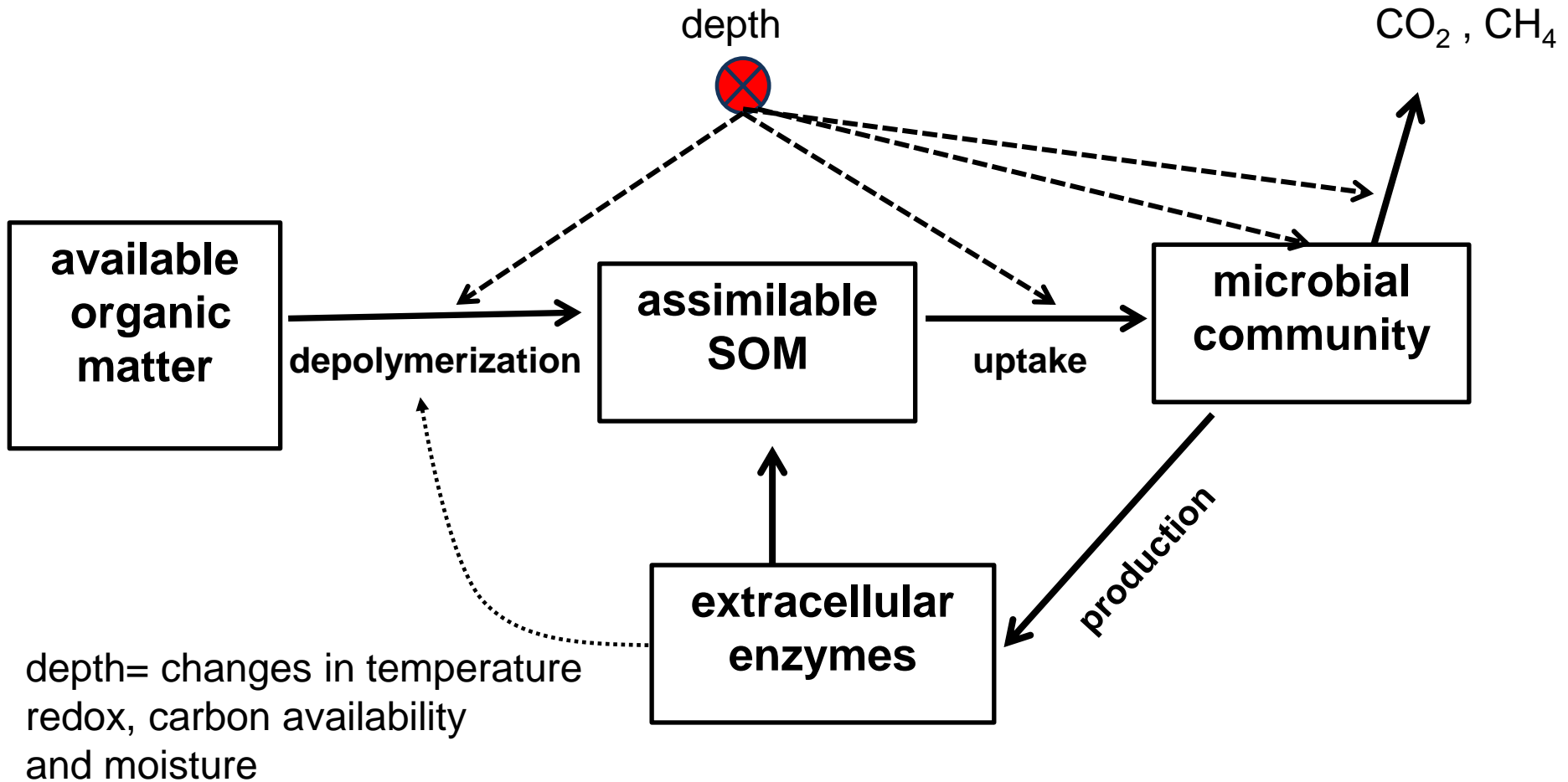
- **Poster #333, Session 2: Colleen Iversen**, “Spruce-Peatland Responses Under Climatic and Environmental Change: An In Situ Warming by CO<sub>2</sub> Manipulation of an ombrotrophic bog in northern MN
- <http://mnspruce.ornl.gov/content/doe-office-science-ber-website>

# microbe-centric belowground cycling



simplified from Conant et al. 2011, GCB

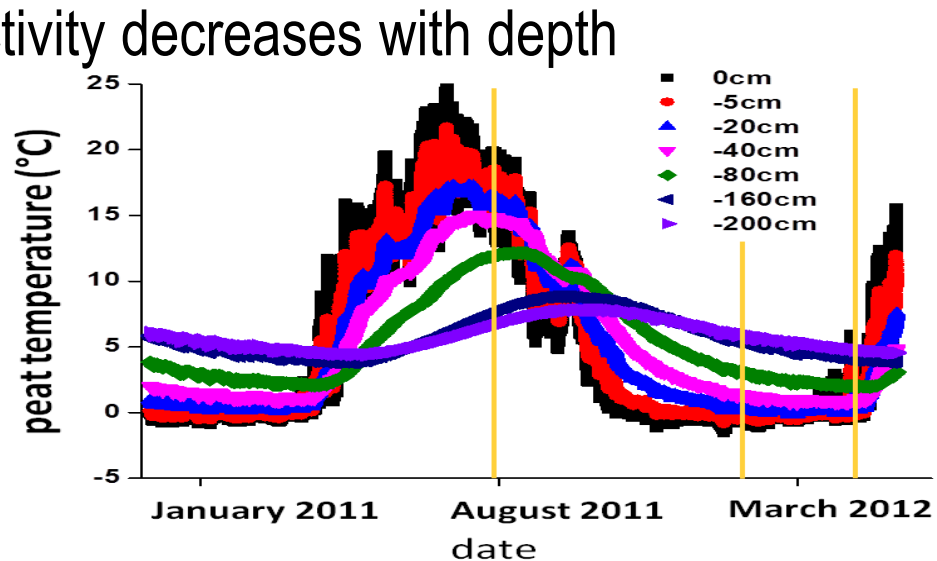
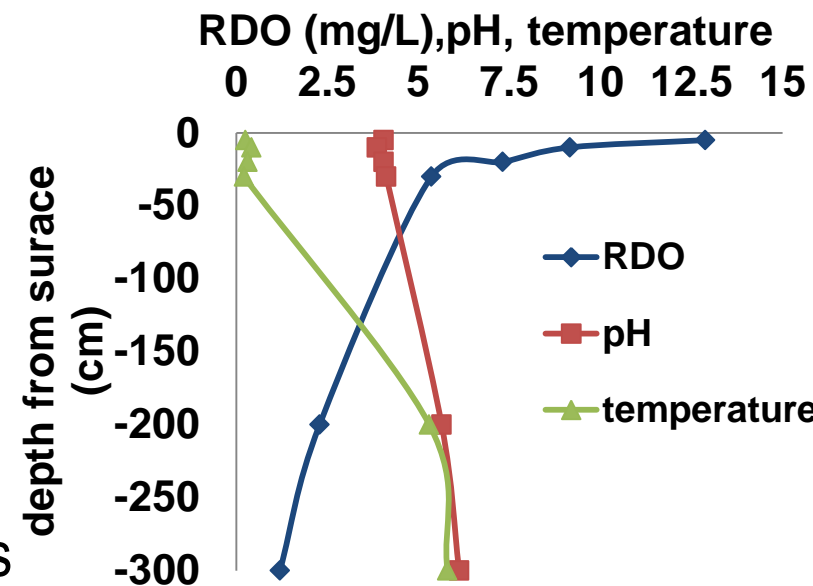
# microbe-centric belowground cycling



simplified from Conant et al. 2011, GCB

# Hypotheses:

- enzyme activity decreases with depth
  - due to reduction in microbial biomass, and/or oxygen content
- temperature response of different enzymes
  - influence N and C depolymerization rates
- temperature response of enzyme activity decreases with depth
  - relatively stable temperature below 1m
  - Isoenzymes, enzymes with same function but different temperature optima, may be more prevalent at the surface.



# methods: sample collection

- peat collected in summer, winter and spring
- peat cores 0-200cm, collected in 50cm increments
  - Divided into 10 or 25cm samples
- stored cold until frozen at  $-20^{\circ}$  C



# methods: potential enzyme activity

peat slurry with sodium acetate buffer, pH 4

mixed with fluorescently labeled substrate

Enzyme Name	Abbreviation	Nutrient Cycle	Enzyme Function
$\beta$ -glucosidase	BG	C	hydrolysis of terminal $\beta$ -glucosyl residues
N-acetyl glucoaminidase	NAG	N	hydrolysis of chitin N-acetyl- $\beta$ -glucosaminide
Cellobiohydrolase	CB	C	hydrolysis of $\beta$ -D-glucosyl linkages
Xylosidase	XYL	C	hydrolysis of $\beta$ -D-xylose residues

incubated at 4, 10, 19 and 30° C

fluorescence measured after 2, 4 and 7 hours

activity expressed as nmol activity g<sup>-1</sup> dry peat hr<sup>-1</sup>



# results: potential enzyme activity with depth

a. August 2011

nmol activity g<sup>-1</sup> dry peat h<sup>-1</sup>

0 1000 2000 3000 4000

b. February 2012

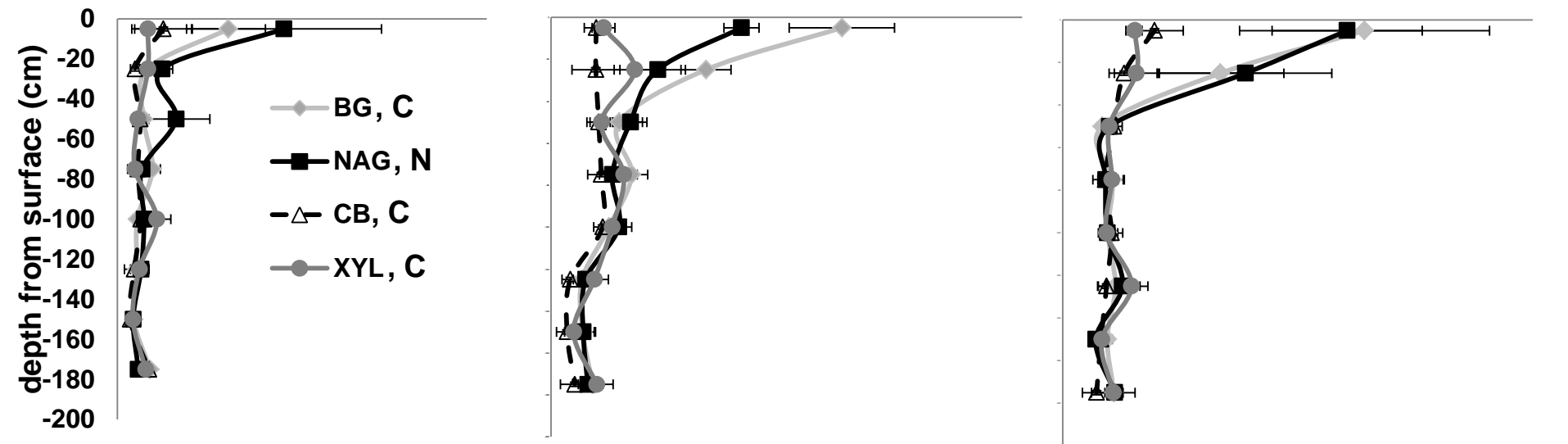
nmol activity g<sup>-1</sup> dry peat h<sup>-1</sup>

0 2000 4000 6000 0

c. April 2012

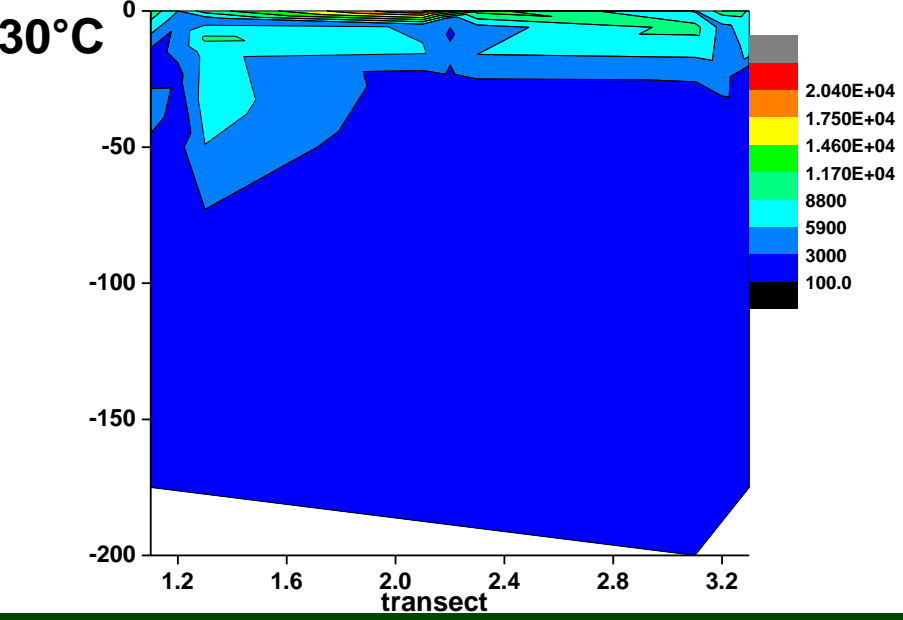
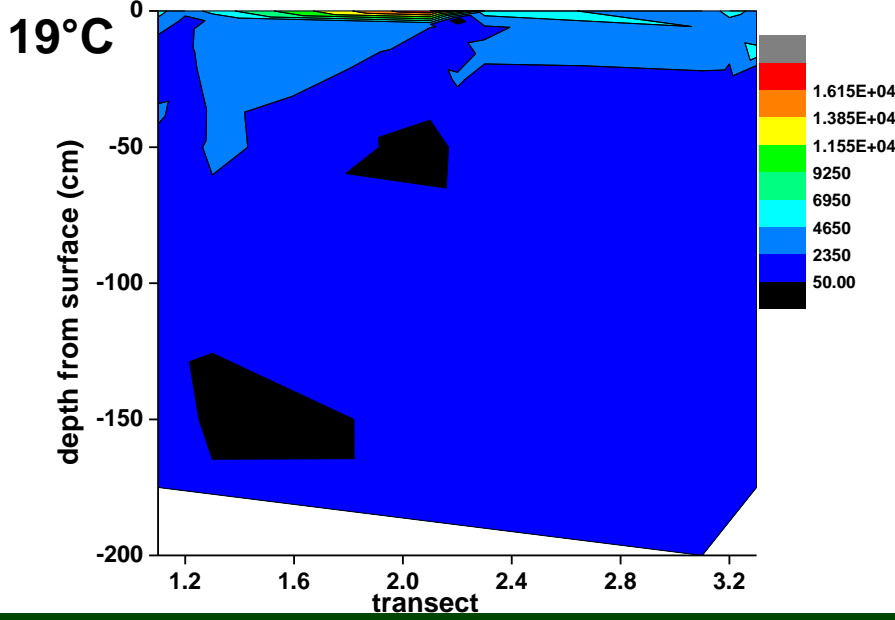
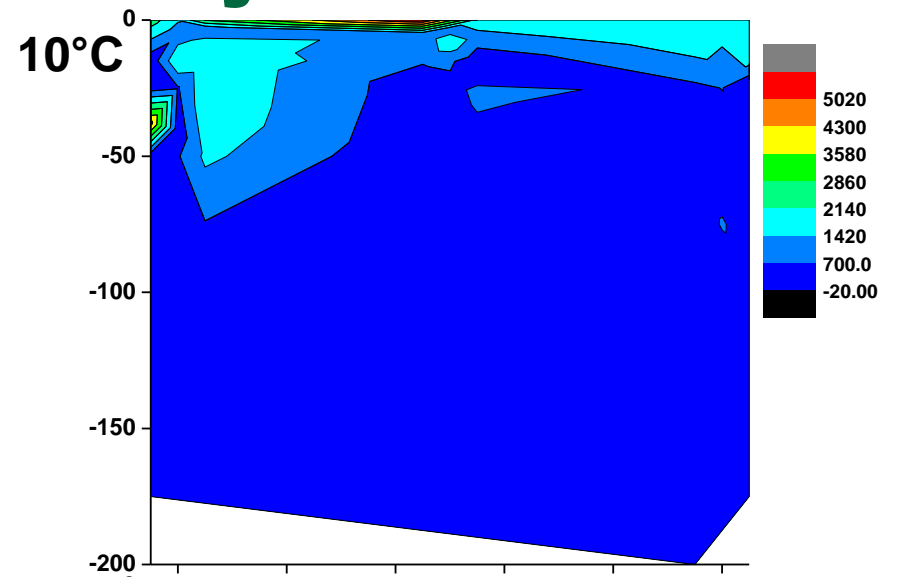
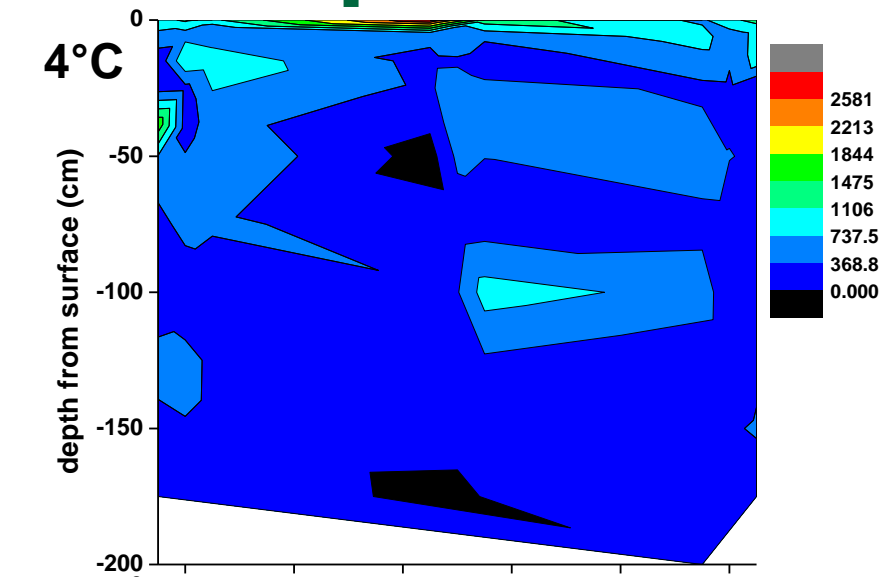
nmol activity g<sup>-1</sup> dry peat h<sup>-1</sup>

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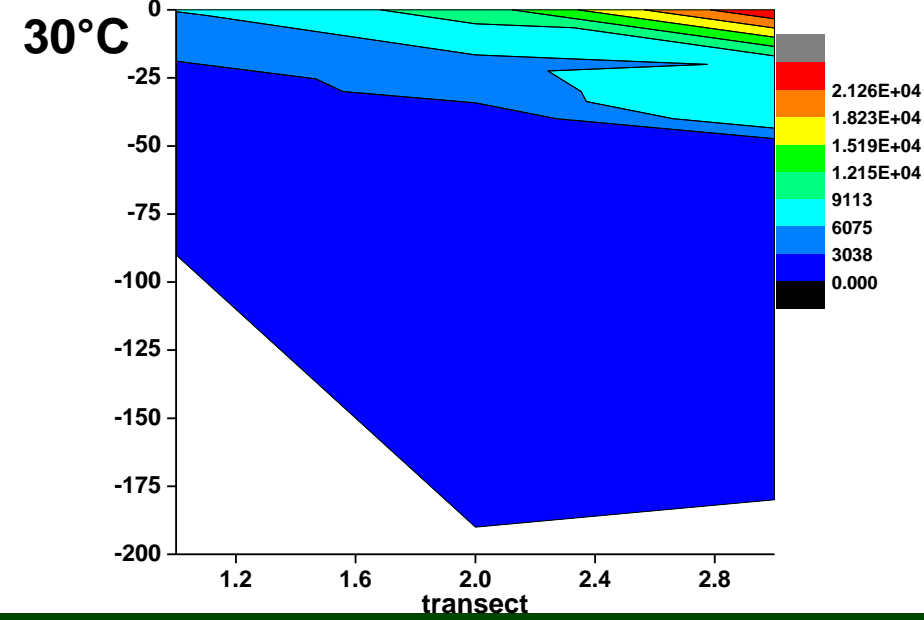
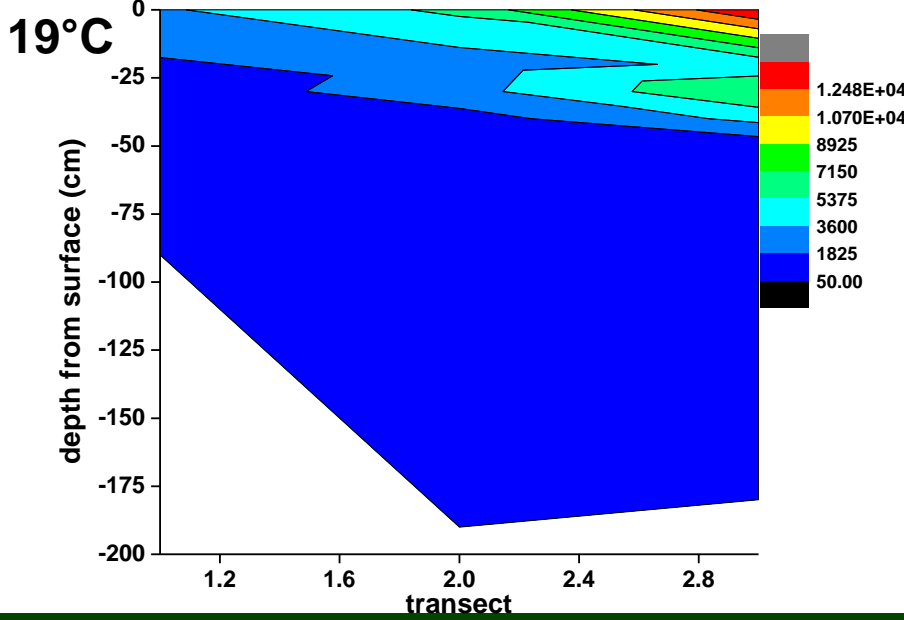
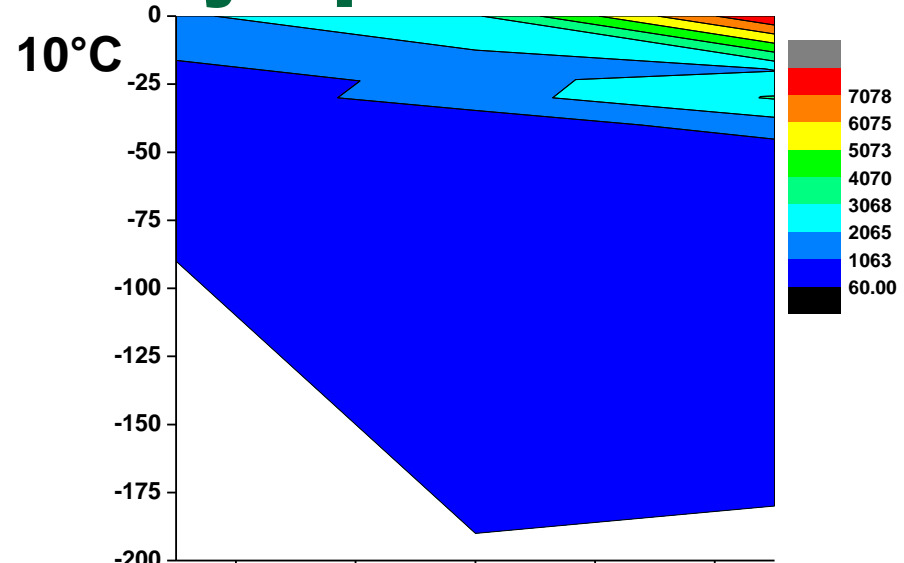
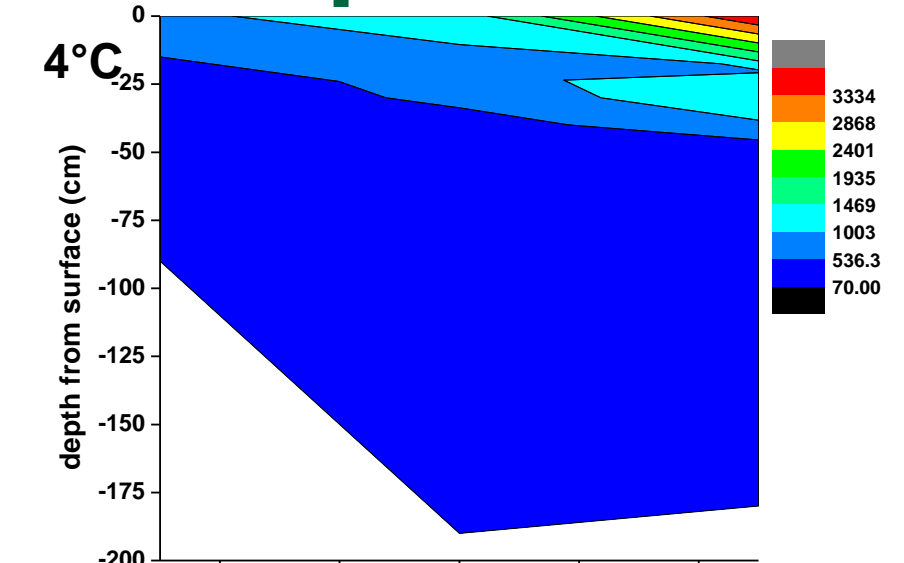


- variability greatest at the surface
- greatest activity in April 2012
- similar trends for all temperatures

# results: potential BG activity Feb 2012

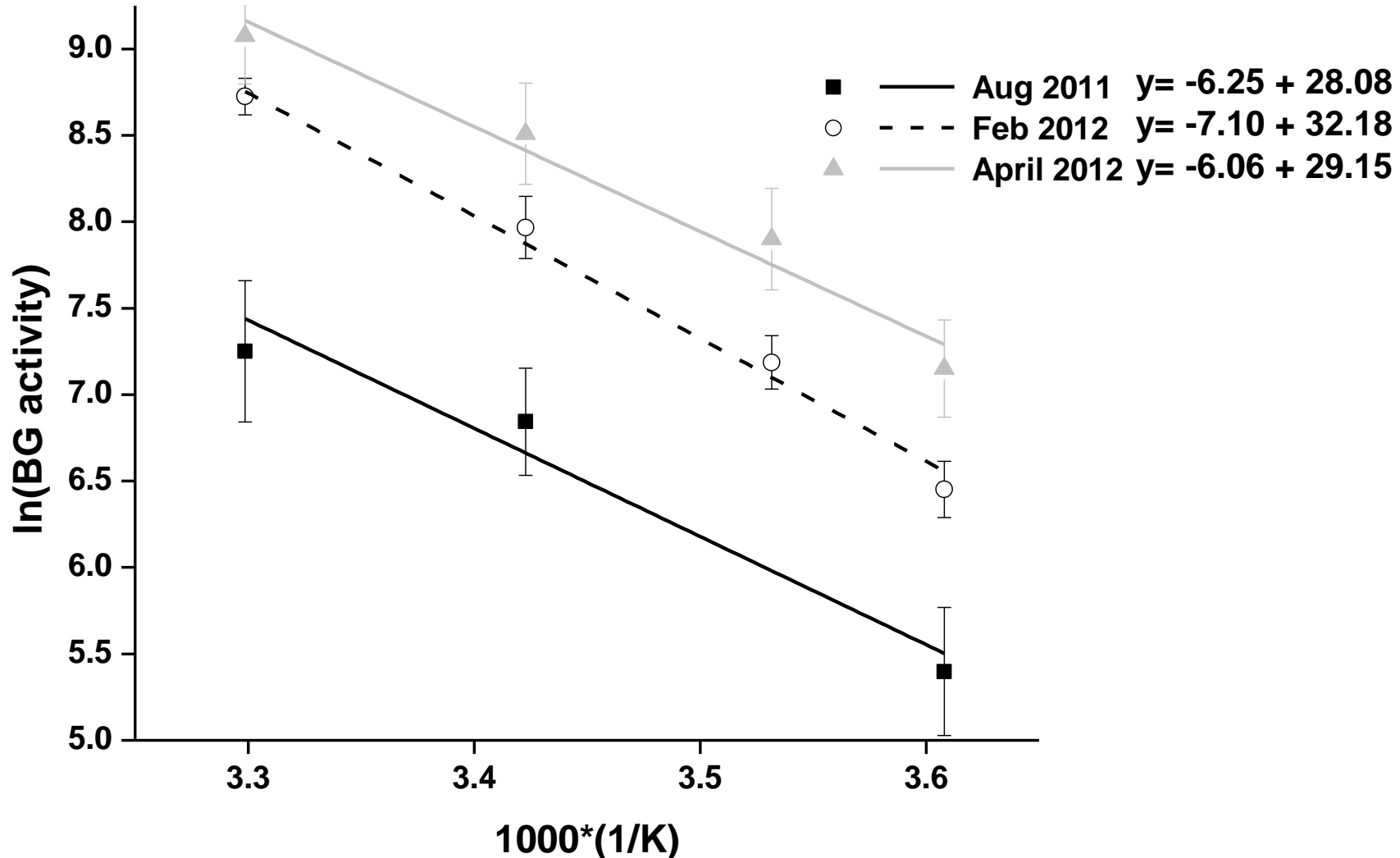


# results: potential BG activity April 2012



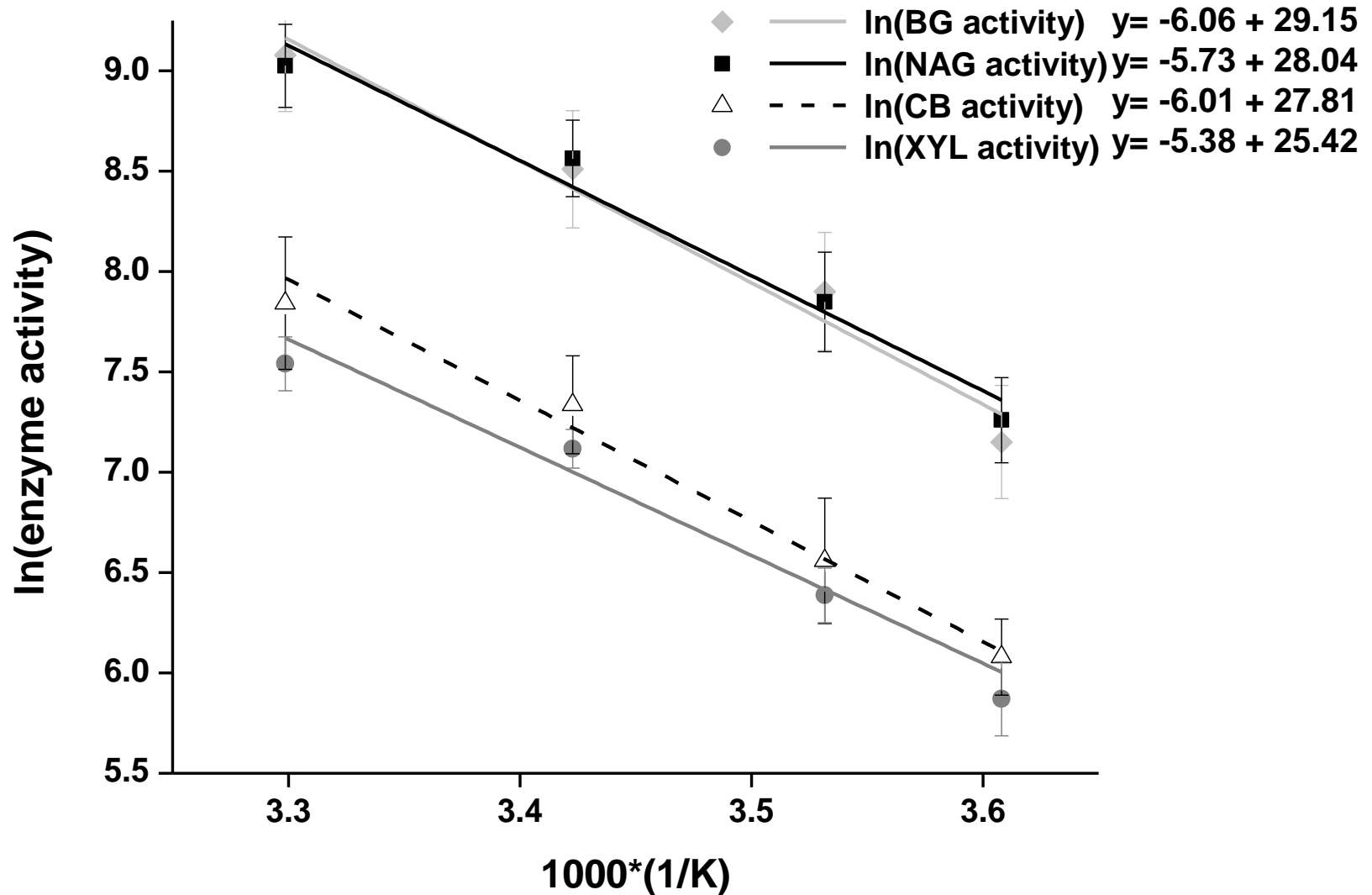
# results: BG temperature response by season

BG activity, all sample dates, 0-25cm below surface



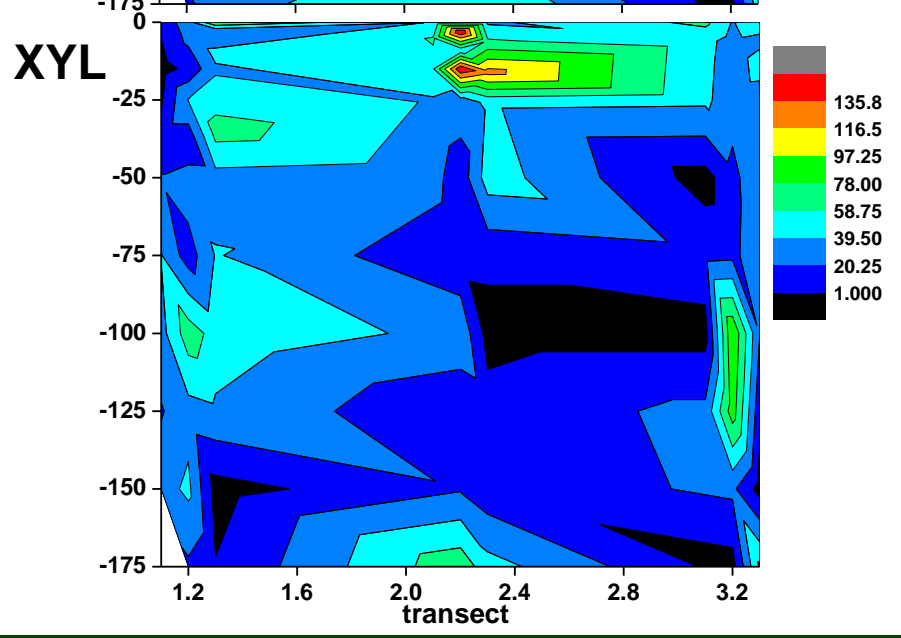
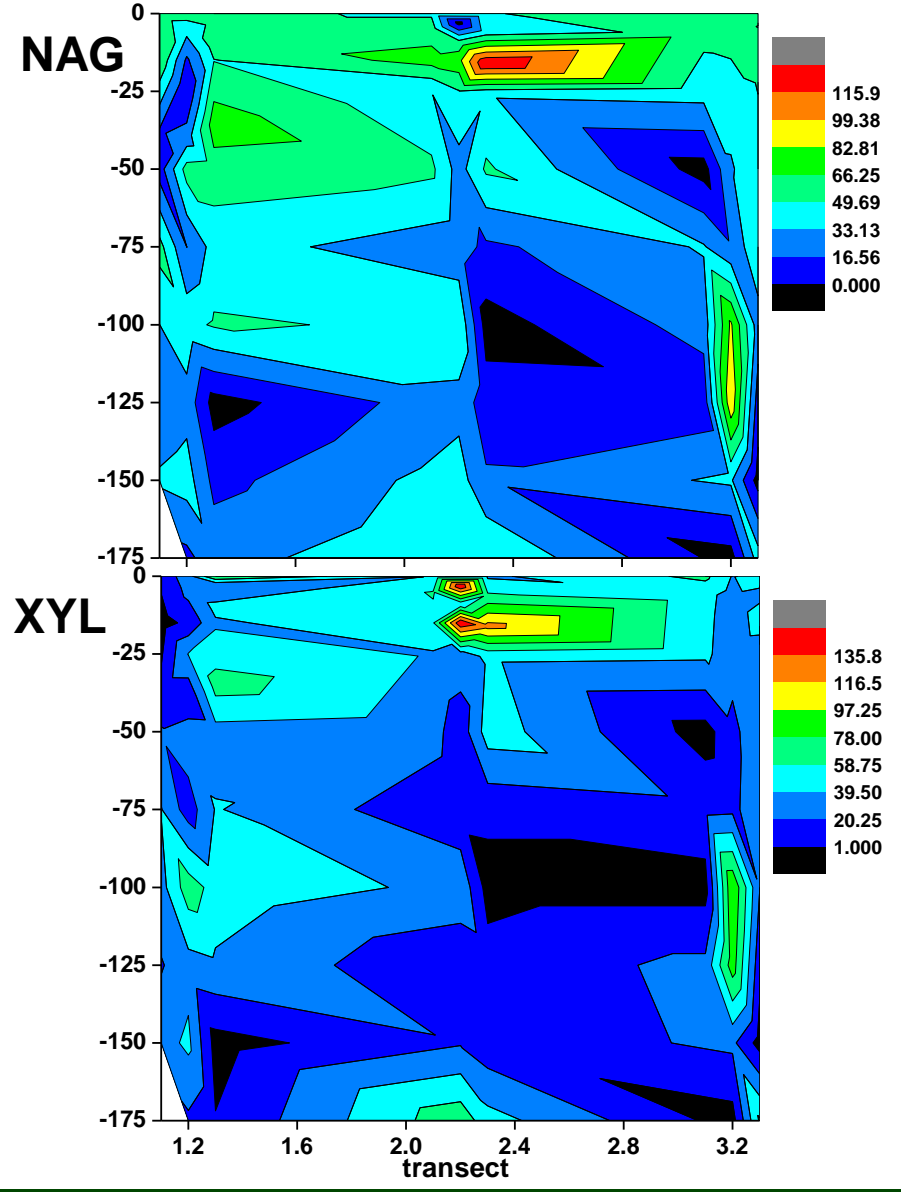
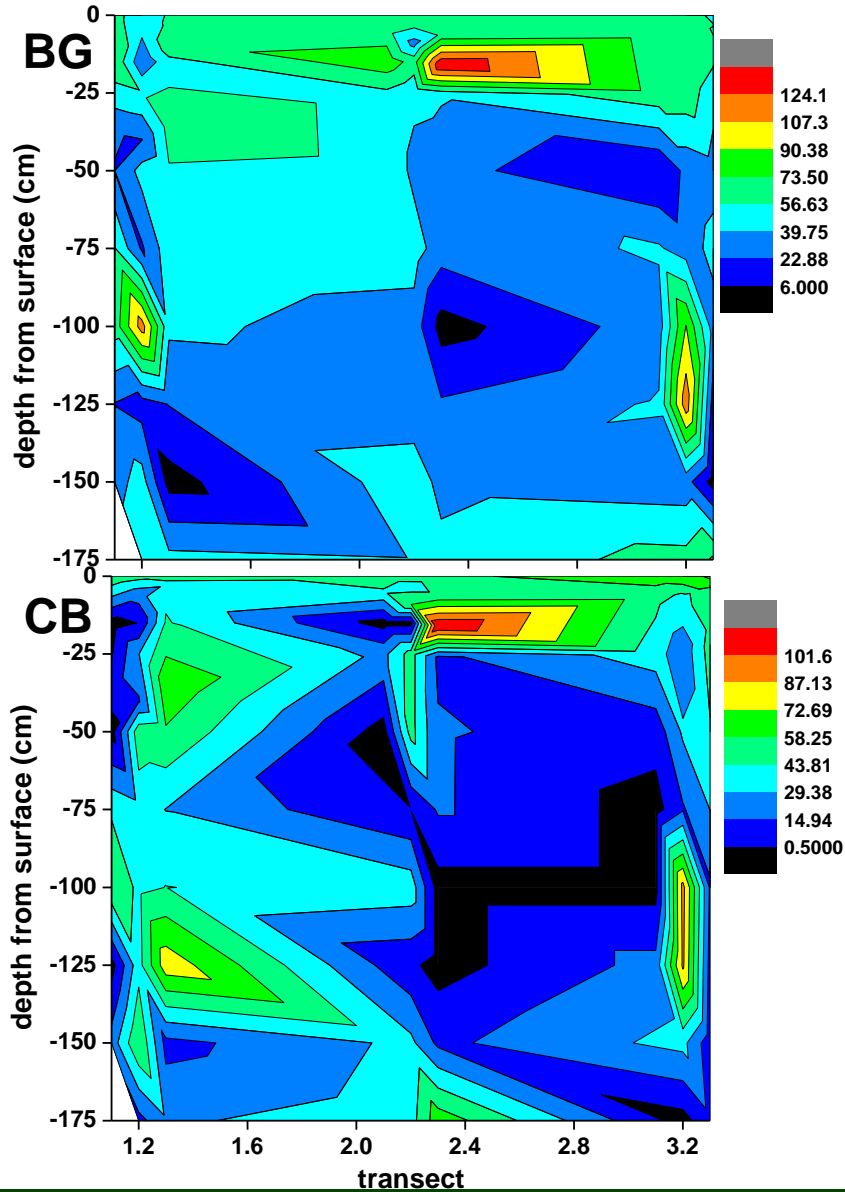
# results: enzyme temperature responses

April 2012, 0-25cm below surface

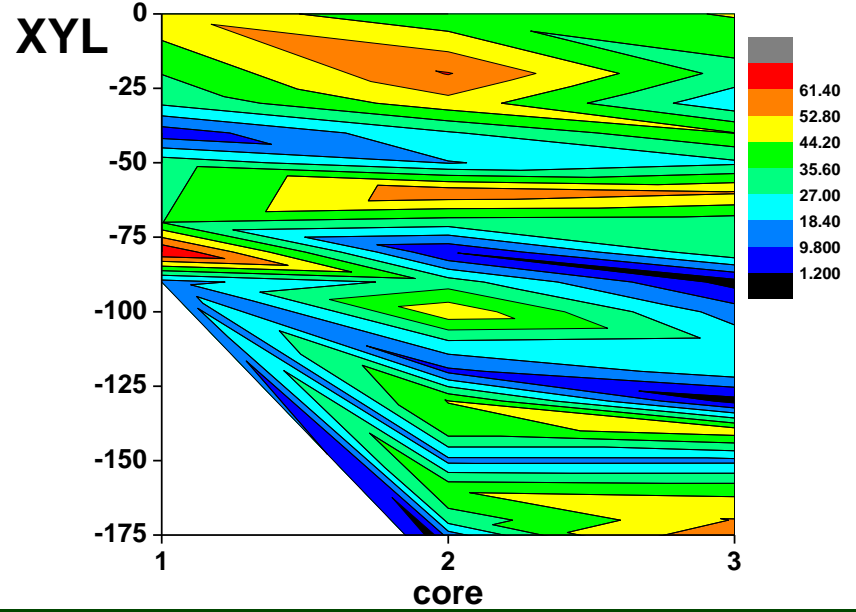
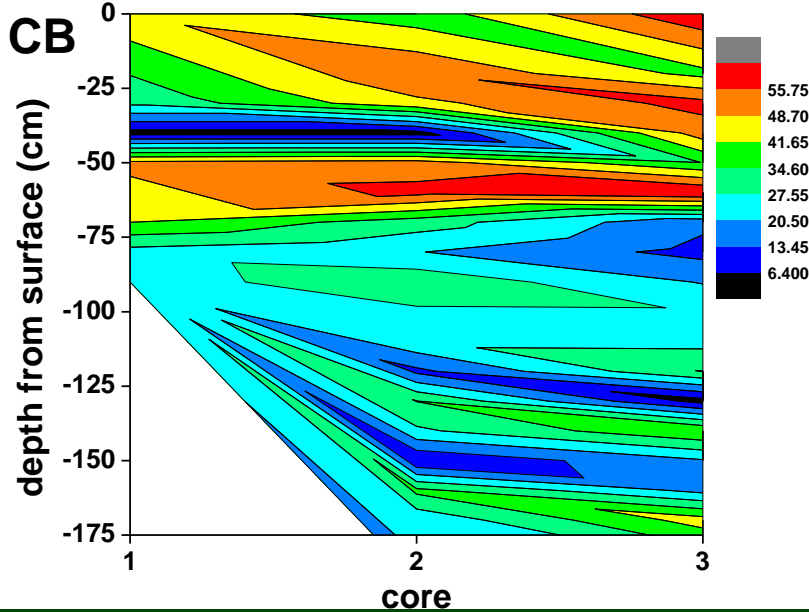
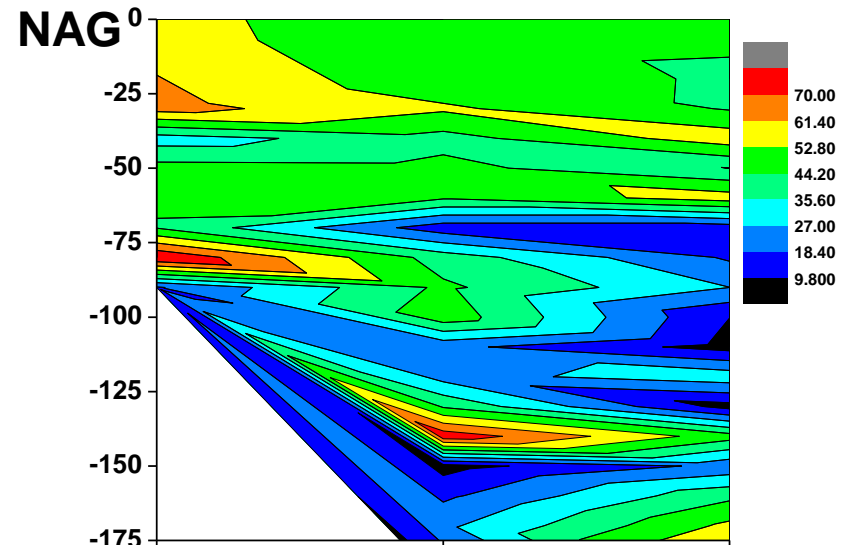
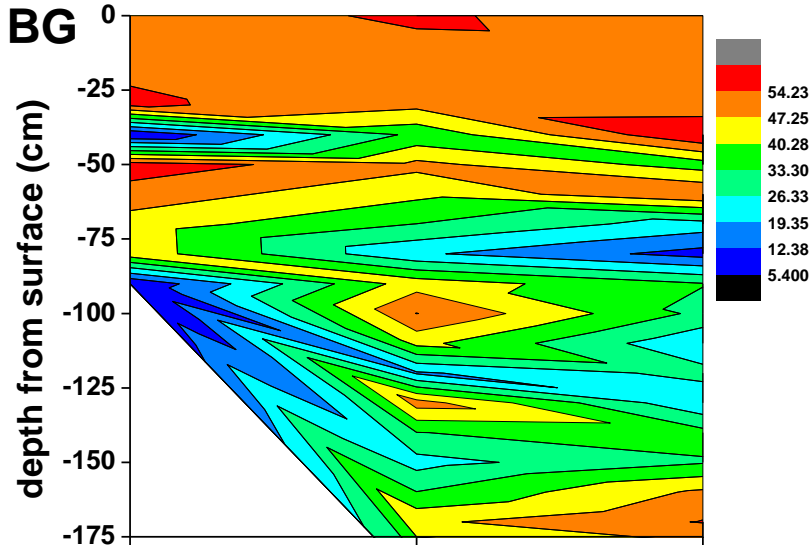




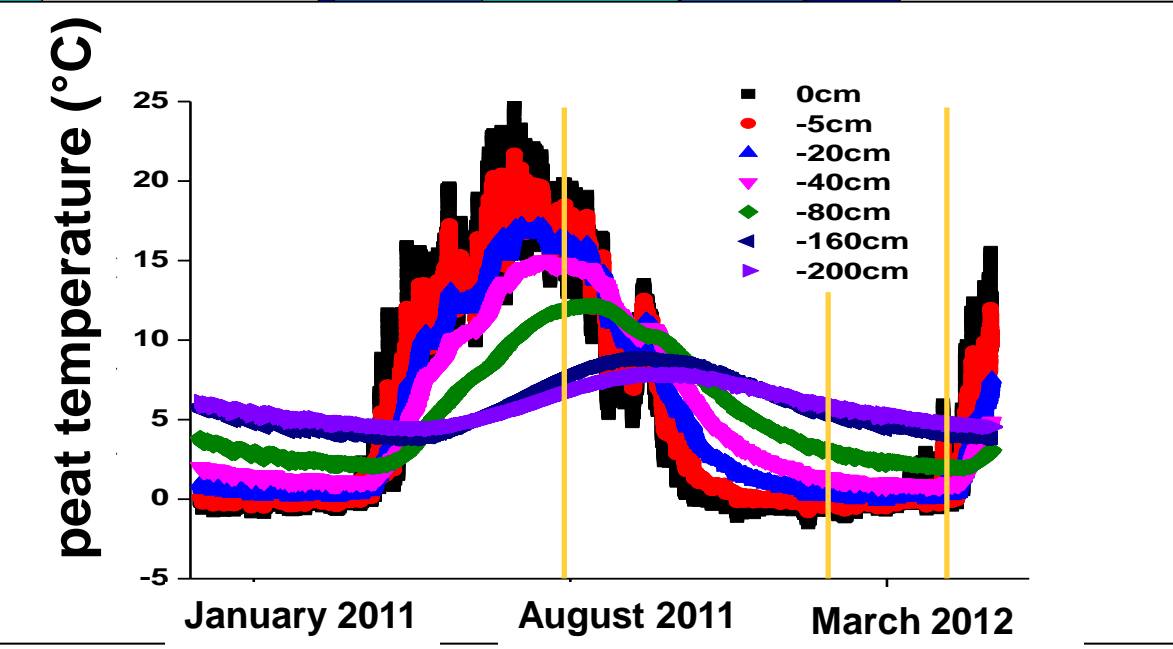
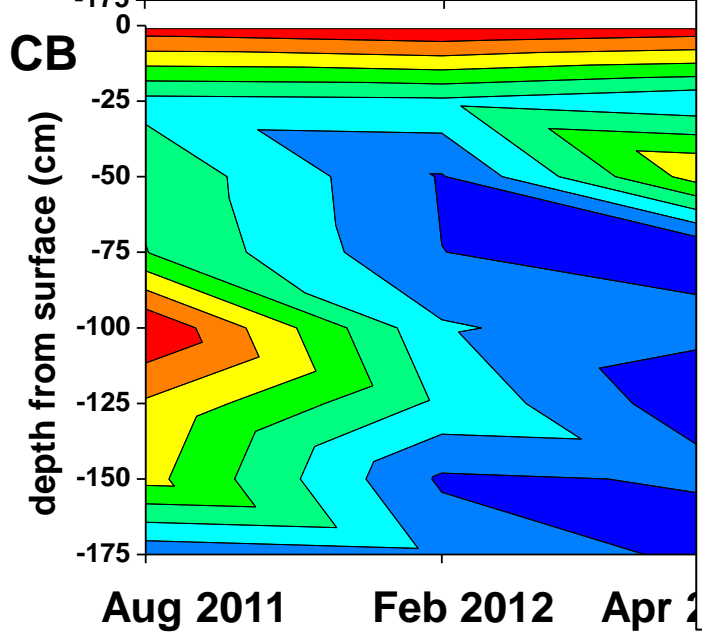
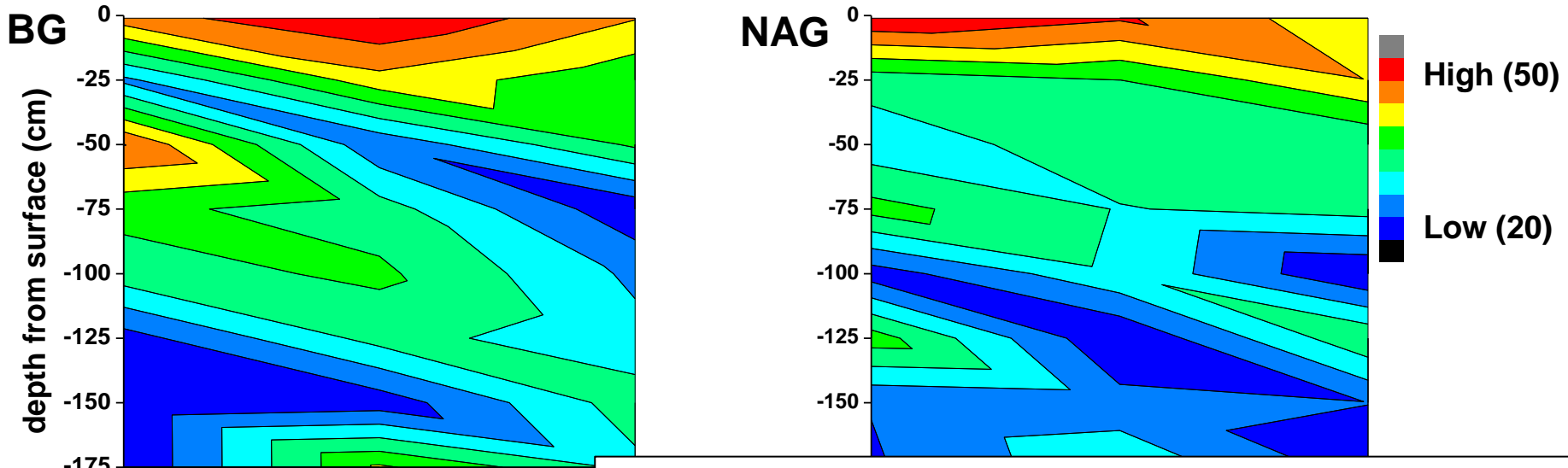
# results: Feb 2012 temperature response by core



# results: April 2012 temperature response by core



# results: average enzyme temperature response



## summary

- potential enzyme activity
  - spatially and temporally heterogeneous
  - greatest in winter, unlikely *in situ* activity greatest
- temperature response
  - spatially and temporally heterogeneous
  - season transitions

## future work

- include N and P enzymes
- decrease sampling distance
- measure dissolved oxygen, characterize microbial community